




# Draft Genome Sequences of *Pantoea agglomerans*, *Paenibacillus polymyxa*, and *Pseudomonas* sp. Strains, Seed Biogel-Associated Endophytes of *Cucumis sativus* L. (Cucumber) and *Cucumis melo* L. (Cantaloupe)

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**ABSTRACT** We report here the draft genome sequences of strains of *Pantoea agglomerans* (EKM10T, EKM20T, EKM21T, and EKM22T), *Paenibacillus polymyxa* (EKM10P and EKM11P), and *Pseudomonas* sp. strain EKM23D. These microbes were cultured from fresh seed biogels of *Cucumis sativus* L. (cucumber) and *Cucumis melo* L. (cantaloupe). The strains suppress the growth of soilborne fungal/oomycete phytopathogens *in vitro*.

The seeds of many plants are coated with biogels (mucilage) consisting of polysaccharides that have been shown to promote the growth of animal gut microbes (1), but surprisingly little is known as to whether these biogels host native microbes. The cucurbit family, which includes cucumber and cantaloupe, is well known for its seed biogels. We previously showed that the biogels (mucilage) coating the seeds of different cucurbits host microbes with antimicrobial protective functions (2). This is analogous to defensive antimicrobial biomolecules in the amniotic fluid surrounding developing animal embryos (3, 4). We previously explored the genomes of biogel microbes from wild cucumber (*Echinocystis lobata*) (5). As such research is novel in the literature, the rationale of this study is to further explore native seed biogel microbe genomes, here from fresh *Cucumis sativus* L. (cucumber) and *Cucumis melo* L. (cantaloupe) fruits.

The fruits were grown on commercial farms in Ontario, Canada, and purchased from local markets in the city of Guelph. Following sterilization of the fruit (which was washed with water and soap, sprayed with 70% ethanol, and allowed to dry), the seeds of three fresh fruits (3 replicates) of each crop were aseptically extracted, including their surrounding biogels. The seeds were transferred into Falcon tubes and washed three times with autoclaved double-distilled water (ddH<sub>2</sub>O); then, 100  $\mu$ l of each wash was streaked onto three agar media (LGI [5], potato dextrose agar [PDA], and Reasoner's 2A [R2A] agar) (6). From the two crops, here we focus on seven unique bacterial colonies that were isolated and identified using the 16S gene primer pair 799F and 1492R by performing a BLAST search against the NCBI and RDP databases. Three strains originated from the cucumber (*Pantoea* sp. strain EKM10T, *Paenibacillus* sp. strains EKM10P and EKM11P) and four strains from the cantaloupe (*Pantoea* sp. strains EKM20T, EKM21T, and EKM22T and *Pseudomonas* sp. strain EKM23D); they were deposited in GenBank (under the accession numbers MK852351.1, MK852340.1, MK852345.1, MK852348.1, MK852349.1, MK852358.1, MK852361.1, respectively). Several strains of *Pantoea*, *Paenibacillus*, and *Pseudomonas* have been registered/commercialized as crop biocontrol agents (7–10); thus, we previously tested these microbes *in vitro* for their suppressive potential against soilborne pathogens. All strains suppressed oomycetes (*Phytophthora capsici* and *Pythium aphanidermatum*), but *Paenibacillus* sp. strains

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**TABLE 1** Characteristics and accession numbers of sequenced genomes of bacterial endophytes isolated from fresh cucumber and cantaloupe seed biogels

Isolate	Bacterial species <sup>a</sup>	Genome size (bp)	No. of clean reads	No. of contigs	$N_{50}$ (bp)	No. of genes	G+C content (%)	SRA accession no.	GenBank accession no.
EKM10T	<i>Pantoea agglomerans</i>	4,710,579	2,402,254	55	387,357	4,064	55	SRR11051778	JAALEU000000000.1
EKM20T	<i>Pantoea agglomerans</i>	4,886,663	2,311,352	69	390,007	4,180	56	SRR11051698	JAALFW000000000.1
EKM21T	<i>Pantoea agglomerans</i>	4,874,245	2,445,781	39	420,936	4,170	55	SRR11051719	JAALFY000000000.1
EKM22T	<i>Pantoea agglomerans</i>	4,875,195	2,370,526	50	380,273	4,172	55	SRR11051699	JAALFX000000000.1
EKM10P	<i>Paenibacillus polymyxa</i>	5,911,943	1,759,810	178	468,459	4,650	52	SRR11038269	JAALET000000000.1
EKM11P	<i>Paenibacillus polymyxa</i>	5,909,556	1,420,012	185	670,170	4,663	51	SRR11051718	JAALEV000000000.1
EKM23D	<i>Pseudomonas</i> sp.	5,796,589	2,460,061	155	197,140	4,891	60	SRR11051781	JAALFY000000000.1

<sup>a</sup> Taxonomy of sequenced strains was collected from updated GenBank databases.

EKM10P and EKM11P exclusively inhibited fungal growth (*Fusarium graminearum* and *Rhizoctonia solani*) (2).

Unique strains were preserved and maintained as 50% glycerol stocks at  $-80^{\circ}\text{C}$ , which were used for all downstream experiments. From the original glycerol stocks, strains were streaked onto LB agar and incubated overnight at  $30^{\circ}\text{C}$ . Single colonies were selected to inoculate LB broth and were incubated overnight in an orbital shaker at  $37^{\circ}\text{C}$  and 250 rpm; then, genomic DNA was extracted from the harvested bacterial pellets using DNeasy UltraClean microbial kits (12224-50; Qiagen), adjusted to  $50\text{ ng}/\mu\text{l}$ . Libraries were prepared using TruSeq DNA Nano library prep kits (KAPA HyperPrep kit, kit code KK8504) and then sequenced using the Illumina NovaSeq 6000 platform. The numbers of output raw reads ( $2 \times 150\text{-bp}$  paired-end format) were 3,149,983 (EKM10T), 3,144,054 (EKM20T), 3,373,222 (EKM21T), 3,170,502 (EKM22T), 2,222,688 (EKM10P), 1,748,173 (EKM11P), and 3,380,471 (EKM23D). The EvoCAT pipeline (Evogene Clustering & Assembly Toolbox) was used to perform *de novo* assembly. The taxonomy was identified using KmerFinder 3.1 (11), with  $178\times$  (EKM10T),  $172\times$  (EKM20T),  $182\times$  (EKM21T),  $176\times$  (EKM22T),  $91\times$  (EKM10P),  $74\times$  (EKM11P), and  $105\times$  (EKM23D) sequence coverages when a BLAST search was performed against the top sequence matches of *Pantoea agglomerans* strain L15 (GenBank accession number [NZ\\_CP034148](#)), *Paenibacillus polymyxa* strain HY96-2 ([NZ\\_CP025957](#)), and *Pseudomonas* sp. strain TKP ([CP006852](#)), with 67.66%, 65.12%, 65.04%, 65.03%, 75.29%, 75.33%, and 14.7% query coverages, respectively. Protein predictions were generated using Prodigal software (12); then, the NCBI nonredundant protein database was queried using BLASTP software to identify the most similar sequences (13). Peptide domains were determined using InterProScan 5.32-71.0 (14). Unless otherwise specified, default settings were used for all software. The characteristics and accession numbers of the genome sequences are presented in Table 1.

All strains possessed genes predicted to encode biomolecules potentially involved in their previously identified antagonistic activities, including bacteriocins (antimicrobial peptides) (15), hydrolytic enzymes (e.g., chitinases [16], proteases [17], pectin/pectate lyase [18]), siderophore-like molecules (19, 20), polyketide synthase (PKS) (8, 21), phenazine biosynthesis protein (PhzF) (19, 22, 23), and metalloenzyme and LuxS/M16 peptidase-like (involved in quorum sensing) (24, 25). Noteworthy is that the *Paenibacillus* strains lacked PhzF but uniquely carried genes implicated in the biosynthesis of  $\beta$ -glucanase (16, 26) and lantibiotics (27). These strains along with *Pseudomonas* sp. strain EKM23D encode cellulases (28, 29) and subtilisin serine protease (30). These findings implicate a diversity of genetic mechanisms underlying the biocontrol activities of seed biogels, a novel microbiome ecological niche.

**Data availability.** This whole-genome shotgun project and raw Illumina reads have been deposited in DDBJ/EMBL/GenBank and the SRA, respectively, under the accession numbers noted in Table 1.

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